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PATENT APPLICATION

TITLE

HUMANIZED ANTIBODIES AGAINST LEUKOCYTE ADHESION MOLECULE VLA-4

Inventors: MARY M. BENDIG, a citizen of USA, residing at 53 Solent Road, West Hampstead, London, NW6 1TX, United Kingdom;

OLIVIER J. LÉGER, a citizen of France, residing at 24 Moxon Street Barnet EN5 5TS, Hertfordshire, United Kingdom;

JOSÉ SALDANHA, a citizen of UK, residing at 22A Lincoln Way, Enfield, Middlesex, EN1 1TE, United Kingdom; and

S. TARRAN JONES, a citizen of UK, residing at 10 The Close, Radlett, Hertfordshire WD7 8HA, United Kingdom.

Assignees: ATHENA NEUROSCIENCES, INC.

and

MRC COLLABORATIVE CENTRE

Status: Small Entity

TOWNSEND and TOWNSEND KHOURIE and CREW
Steuart Street Tower, 20th Floor
One Market Plaza
San Francisco, California 94105
(415) 543-9600

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TECHNICAL FIELD

This invention relates generally to humanized antibodies specific for the alpha-4 subunit of leukocyte adhesion molecule VLA-4.

BACKGROUND OF THE INVENTION

Inflammation is a response of vascularized tissues to infection or injury and is effected by adhesion of leukocytes to the endothelial cells of blood vessels and their infiltration into the surrounding tissues. In normal inflammation, the infiltrating leukocytes release toxic mediators to kill invading organisms, phagocytize debris and dead cells, and play a role in tissue repair and the immune response. However, in pathologic inflammation, infiltrating leukocytes are over-responsive and can cause serious or fatal damage. *See, e.g.,* Hickey, *Psychoneuroimmunology II* (Academic Press 1990).

The attachment of leukocytes to endothelial cells is effected via specific interaction of cell-surface ligands and receptors on endothelial cells and leukocytes. *See generally* Springer, *Nature* 346:425-433 (1990). The identity of the ligands and receptors varies for different cell subtypes, anatomical locations and inflammatory stimuli. The VLA-4 leukocyte cell-surface receptor was first identified by Hemler, EP 330,506 (1989) (incorporated by reference in its entirety for all purposes). VLA-4 is a member of the $\beta 1$ integrin family of cell surface receptors, each of which comprises α and β chains. VLA-4 contains an $\alpha 4$ chain and a $\beta 1$ chain. VLA-4 specifically binds to an endothelial cell ligand termed VCAM-1. *See* Elices et al., *Cell* 60:577-584 (1990) (incorporated by reference in its entirety for all purposes). Although VCAM-1 was first detected on activated human umbilical vein cells, this ligand has also been detected on brain endothelial cells.

See commonly owned, co-pending application US Serial No. 07/871,223 (incorporated by reference in its entirety for all purposes).

Adhesion molecules such as VLA-4, are potential targets for therapeutic agents. The VLA-4 receptor is a particularly important target because of its interaction with a ligand residing on brain endothelial cells. Diseases and conditions resulting from brain inflammation have particularly severe consequences. For example, one such disease, multiple sclerosis (MS), has a chronic course (with or without exacerbations and remissions) leading to severe disability and death. The disease affects an estimated 250,000 to 350,000 people in the United States alone.

Antibodies against the VLA-4 receptor have been tested for their anti-inflammatory potential both *in vitro* and *in vivo* in animal models. *See* USSN 07/871,223 and Yednock et al., *Nature* 356:63-66 (1992) (incorporated by reference in its entirety for all purposes). The *in vitro* experiments demonstrate that anti-VLA-4 antibodies block attachment of lymphocytes to brain endothelial cells. The animal experiments test the effect of anti-VLA-4 antibodies on animals having an artificially induced condition (experimental autoimmune encephalomyelitis), simulating multiple sclerosis. The experiments show that administration of anti-VLA-4 antibodies prevents inflammation of the brain and subsequent paralysis in the animals. Collectively, these experiments identify anti-VLA-4 antibodies as potentially useful therapeutic agents for treating multiple sclerosis and other inflammatory diseases and disorders.

A significant problem with the anti-VLA-4 antibodies available to-date is that they are all of murine origin, and therefore likely to raise a human anti-mouse response (HAMA) in clinical use. A HAMA response reduces the efficacy of mouse antibodies in patients and prevents continued administration. One approach to this problem is to humanize mouse antibodies. In this approach, complementarity determining regions (CDRs) and certain other amino acids from donor mouse variable regions are grafted into human variable acceptor regions and then

joined to human constant regions. *See, e.g.*, Riechmann et al., *Nature* 332:323-327 (1988); Winter, US 5,225,539 (1993) (each of which is incorporated by reference in its entirety for all purposes).

Although several examples of humanized antibodies have been produced, the transition from a murine to a humanized antibody involves a compromise of competing considerations, the solution of which varies with different antibodies. To minimize immunogenicity, the immunoglobulin should retain as much of the human acceptor sequence as possible. However, to retain authentic binding properties, the immunoglobulin framework should contain sufficient substitutions of the human acceptor sequence to ensure a three-dimensional conformation of CDR regions as close as possible to that in the original mouse donor immunoglobulin. As a result of these competing considerations, many humanized antibodies produced to-date show some loss of binding affinity compared with the corresponding murine antibodies from which they are derived. *See, e.g.*, Jones et al., *Nature* 321:522-525 (1986); Shearman et al., *J. Immunol.* 147:4366-4373 (1991); Kettleborough et al., *Protein Engineering* 4:773-783 (1991); Gorman et al., *Proc. Natl. Acad. Sci. USA* 88:4181-4185 (1991); Tempest et al., *Biotechnology* 9:266-271 (1991).

Based on the foregoing it is apparent that a need exists for humanized anti-VLA-4 antibodies demonstrating a strong affinity for the VLA-4 receptor, while exhibiting little, if any, human-antimouse response. The present invention fulfill this and other needs.

SUMMARY OF THE INVENTION

The invention provides humanized immunoglobulins that specifically bind to a VLA-4 ligand. The humanized antibodies comprise a humanized light chain and a humanized heavy chain. The humanized light chain comprises three complementarity determining regions (CDR1, CDR2 and CDR3) having amino acid sequences from the corresponding complementarity determining regions of a mouse 21-6 immunoglobulin light chain, and a variable region framework from a human kappa light chain

variable region framework sequence except in at least one position selected from a first group consisting of positions L45, L49, L58 and L69, wherein the amino acid position is occupied by the same amino acid present in the equivalent position of the mouse 21.6 immunoglobulin light chain variable region framework. The humanized heavy chain comprises three complementarity determining regions (CDR1, CDR2 and CDR3) having amino acid sequences from the corresponding complementarity determining regions of a mouse 21-6 immunoglobulin heavy chain, and a variable region framework from a human heavy chain variable region framework sequence except in at least one position selected from a group consisting of H27, H28, H29, H30, H44, H71, wherein the amino acid position is occupied by the same amino acid present in the equivalent position of the mouse 21-6 immunoglobulin heavy chain variable region framework. The immunoglobulins specifically bind to VLA-4 with an affinity having a lower limit of about 10^7 M^{-1} and an upper limit of about five times the affinity of the mouse 21-6 immunoglobulin.

Usually, the humanized light and heavy chain variable region frameworks are from RE1 and 21/28'CL variable region framework sequences respectively. When the humanized light chain variable region framework is from RE1, at least two framework amino acids are replaced. One amino acid is from the first group of positions described *supra*. The other amino acids is from a third group consisting of positions L104, L105 and L107. This position is occupied by the same amino acid present in the equivalent position of a kappa light chain from a human immunoglobulin other than RE1.

Some humanized immunoglobulins have a mature light chain variable region sequence designated La or Lb in Figure 6, or a mature heavy chain variable region sequence designated Ha, Hb or Hc in Figure 7. Preferred humanized immunoglobulins include those having an La light chain and an Ha, Hb or Hc heavy chain.

The invention also provides binding fragments of the humanized immunoglobulins against VLA-4 described *supra*.

In another aspect, the invention provides nucleic acids encoding the humanized immunoglobulins against VLA-4 described *supra*.

Also provided are computers programmed to display three dimensional images of the mouse 21.6 antibody or the humanized immunoglobulins described *supra*.

In another aspect the invention provides pharmaceutical compositions and methods of treatment using the same. The pharmaceutical compositions comprise a humanized immunoglobulin or binding fragment as described *supra*, and a pharmaceutically acceptable carrier. In some methods of treatment a therapeutically effective amount of a pharmaceutical composition is administered to a patient suffering from an inflammatory disease, such as multiple sclerosis.

Also provided are methods of detecting VLA-4 antigen using the humanized immunoglobulins and binding fragments described *supra*. In these methods, a humanized antibody or binding fragment is administered to a patient or a tissue sample therefrom. Complexes formed by specific binding between the antibody or fragment and VLA-4 present in the sample are detected.

BRIEF DESCRIPTION OF FIGURES

Figure 1: DNA (SEQ ID NO:1) and amino acid (SEQ ID NO:2) sequences of the mouse 21.6 light chain variable region.

Figure 2: DNA (SEQ ID NO:3) and amino acid (SEQ ID NO:4) sequences of the mouse 21.6 heavy chain variable region.

Figure 3: Light (A) and heavy (B) chain expression vectors used to produce chimeric and reshaped human antibodies with human kappa light chains and human gamma-1 heavy chains in mammalian cells.

Figure 4: ELISA comparison of chimeric and mouse 21.6 antibody binding to L cells expressing human $\alpha 4\beta 1$ integrin on their surface.

Figure 5: Molecular model of the variable regions of mouse 21.6 antibody. Residues of special interest are labelled.

Figure 6. Comparisons of the amino acid sequences of mouse and reshaped human 21.6 light chain variable regions. (SEQ ID NO:15) The amino acid residues that are part of the Chothia canonical sequences for the CDR loop structures are marked with an asterisk. REI shows the FRs and CDRs from the V_L region of human REI light chain. (SEQ ID NO:6) La and Lb are the two versions of reshaped human 21.6 V_L region. (SEQ ID NO:7) (SEQ ID NO:8) The residues in the FRs of La that differ from those in the REI sequence are underlined. In Lb, only the residues in the framework regions that differ from those of REI are shown.

Figure 7. Comparisons of the amino acid sequences of the mouse and reshaped human 21.6 heavy chain variable regions. (SEQ ID NO:9) The amino acid residues that are part of the canonical sequences for the Chothia CDR loop structures are marked with an asterisk. 2*CL shows the FRs and CDRs from the V_H region of human 21/28'CL antibody. (SEQ ID NO:10) Ha, Hb and Hc are the three versions of reshaped human 21.6 V_H region. (SEQ ID NO:11) (SEQ ID NO:12) (SEQ ID NO:13) The residues in the FRs of Ha that differ from those in the 21/28'CL sequence are underlined. In Hb and Hc, only the residues in the framework regions that differ from those of 21/28'CL are shown.

Figure 8: PCR-based construction of version "a" of reshaped human 21.6 light chain variable region. The dotted lines indicate a complementary sequence of at least 21 bases between the primers.

Figure 9: PCR-based construction of version "a" of reshaped human 21.6 heavy chain variable region.

Figure 10: cDNA and amino acid sequences of the first version ("a") of reshaped human 21.6 light chain variable region. (SEQ ID NOS:14 and 15)

Figure 11: DNA and amino acid sequences of the first version ("a") of reshaped human 21.6 heavy chain variable region. (SEQ ID NOS:16 and 17)

Figure 12: ELISA comparison of chimeric and reshaped human 21.6 antibodies to bind to L cells expressing human $\alpha 4\beta 1$ integrin on their surface.

DEFINITIONS

Abbreviations for the twenty naturally occurring amino acids follow conventional usage (*Immunology - A Synthesis* (2nd ed., E.S. Golub & D.R. Gren, eds., Sinauer Associates, Sunderland, MA, 1991)). Stereoisomers (*e.g.*, D-amino acids) of the twenty conventional amino acids, unnatural amino acids such as α,α-disubstituted amino acids, N-alkyl amino acids, lactic acid, and other unconventional amino acids may also be suitable components for polypeptides of the present invention. Examples of unconventional amino acids include: 4-hydroxyproline, γ-carboxyglutamate, ϵ-N,N,N-trimethyllysine, ϵ-N-acetyllysine, O-phosphoserine, N-acetylserine, N-formylmethionine, 3-methylhistidine, 5-hydroxylysine, ω-N-methylarginine, and other similar amino acids and imino acids (*e.g.*, 4-hydroxyproline). Moreover, amino acids may be modified by glycosylation, phosphorylation and the like.

In the polypeptide notation used herein, the lefthand direction is the amino terminal direction and the righthand direction is the carboxy-terminal direction, in accordance with standard usage and convention. Similarly, unless specified otherwise, the lefthand end of single-stranded polynucleotide sequences is the 5' end; the lefthand direction of double-stranded polynucleotide sequences is referred to as the 5' direction. The direction of 5' to 3' addition of nascent RNA transcripts is referred to as the transcription direction; sequence regions on the DNA strand having the same sequence as the RNA and which are 5' to the 5' end of the RNA transcript are referred to as "upstream sequences"; sequence regions on the DNA strand having the same sequence as the RNA and which are 3' to the 3' end of the RNA transcript are referred to as "downstream sequences".

The phrase "polynucleotide sequence" refers to a single or double-stranded polymer of deoxyribonucleotide or ribonucleotide bases read from the 5' to the 3' end. It includes self-replicating plasmids, infectious polymers of DNA or RNA and non-functional DNA or RNA.

The following terms are used to describe the sequence relationships between two or more polynucleotides: "reference

sequence", "comparison window", "sequence identity", "percentage of sequence identity", and "substantial identity". A "reference sequence" is a defined sequence used as a basis for a sequence comparison; a reference sequence may be a subset of a larger sequence, for example, as a segment of a full-length cDNA or gene sequence given in a sequence listing, such as a polynucleotide sequence of Figs. 1 or 2, or may comprise a complete DNA or gene sequence. Generally, a reference sequence is at least 20 nucleotides in length, frequently at least 25 nucleotides in length, and often at least 50 nucleotides in length. Since two polynucleotides may each (1) comprise a sequence (*i.e.*, a portion of the complete polynucleotide sequence) that is similar between the two polynucleotides, and (2) may further comprise a sequence that is divergent between the two polynucleotides, sequence comparisons between two (or more) polynucleotides are typically performed by comparing sequences of the two polynucleotides over a "comparison window" to identify and compare local regions of sequence similarity. A "comparison window", as used herein, refers to a conceptual segment of at least 20 contiguous nucleotide positions wherein a polynucleotide sequence may be compared to a reference sequence of at least 20 contiguous nucleotides and wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (*i.e.*, gaps) of 20 percent or less as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Optimal alignment of sequences for aligning a comparison window may be conducted by the local homology algorithm of Smith & Waterman, *Adv. Appl. Math.* 2:482 (1981), by the homology alignment algorithm of Needleman & Wunsch, *J. Mol. Biol.* 48:443 (1970), by the search for similarity method of Pearson & Lipman, *Proc. Natl. Acad. Sci. (USA)* 85:2444 (1988) (each of which is incorporated by reference in its entirety for all purposes), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package Release 7.0, Genetics Computer Group, 575 Science Dr., Madison, WI), or by inspection, and the best alignment (*i.e.*, resulting in the

highest percentage of sequence similarity over the comparison window) generated by the various methods is selected. The term "sequence identity" means that two polynucleotide sequences are identical (*i.e.*, on a nucleotide-by-nucleotide basis) over the window of comparison. The term "percentage of sequence identity" is calculated by comparing two optimally aligned sequences over the window of comparison, determining the number of positions at which the identical nucleic acid base (*e.g.*, A, T, C, G, U, or I) occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison (*i.e.*, the window size), and multiplying the result by 100 to yield the percentage of sequence identity. The terms "substantial identity" as used herein denotes a characteristic of a polynucleotide sequence, wherein the polynucleotide comprises a sequence that has at least 85 percent sequence identity, preferably at least 90 to 95 percent sequence identity, more usually at least 99 percent sequence identity as compared to a reference sequence over a comparison window of at least 20 nucleotide positions, frequently over a window of at least 25-50 nucleotides, wherein the percentage of sequence identity is calculated by comparing the reference sequence to the polynucleotide sequence which may include deletions or additions which total 20 percent or less of the reference sequence over the window of comparison. The reference sequence may be a subset of a larger sequence, for example, the sequence shown in Figs. 1 or 2.

As applied to polypeptides, the term "sequence identity" means peptides share identical amino acids at corresponding positions. The term "sequence similarity" means peptides have identical or similar amino acids (*i.e.*, conservative substitutions) at corresponding positions. The term "substantial identity" means that two peptide sequences, when optimally aligned, such as by the programs GAP or BESTFIT using default gap weights, share at least 80 percent sequence identity, preferably at least 90 percent sequence identity, more preferably at least 95 percent sequence identity or more (*e.g.*, 99 percent sequence identity). Preferably, residue

positions which are not identical differ by conservative amino acid substitutions. The term "substantial similarity" means that two peptide sequences share corresponding percentages of sequence similarity.

The term "substantially pure" means an object species is the predominant species present (*i.e.*, on a molar basis it is more abundant than any other individual species in the composition), and preferably a substantially purified fraction is a composition wherein the object species comprises at least about 50 percent (on a molar basis) of all macromolecular species present. Generally, a substantially pure composition will comprise more than about 80 to 90 percent of all macromolecular species present in the composition. Most preferably, the object species is purified to essential homogeneity (contaminant species cannot be detected in the composition by conventional detection methods) wherein the composition consists essentially of a single macromolecular species.

For purposes of classifying amino acids substitutions as conservative or nonconservative, amino acids are grouped as follows: Group I (hydrophobic sidechains): norleucine, met, ala, val, leu, ile; Group II (neutral hydrophilic side chains): cys, ser, thr; Group III (acidic side chains): asp, glu; Group IV (basic side chains): asn, gln, his, lys, arg; Group V (residues influencing chain orientation): gly, pro; and Group VI (aromatic side chains): trp, tyr, phe. Conservative substitutions involve substitutions between amino acids in the same class. Non-conservative substitutions constitute exchanging a member of one of these classes for another.

Amino acids from the variable regions of the mature heavy and light chains of immunoglobulins are designated Hx and Lxx respectively, where x is a number designating the position of an amino acids according to the scheme of Kabat et al., *Sequences of Proteins of Immunological Interest* (National Institutes of Health, Bethesda, MD (1987) and (1991)) (hereinafter collectively referred to as "Kabat et al.," incorporated by reference in their entirety for all purposes). Kabat et al. list many amino acid sequences for antibodies for

each subclass, and list the most commonly occurring amino acid for each residue position in that subclass. Kabat et al. use a method for assigning a residue number to each amino acid in a listed sequence, and this method for assigning residue numbers has become standard in the field. Kabat et al.'s scheme is extendible to other antibodies not included in the compendium by aligning the antibody in question with one of the consensus sequences in Kabat et al. The use of the Kabat et al. numbering system readily identifies amino acids at equivalent positions in different antibodies. For example, an amino acid at the L50 position of a human antibody occupies the equivalence position to an amino acid position L50 of a mouse antibody.

DETAILED DESCRIPTION

I. Humanized Antibodies Specific for VLA-4

In one embodiment of the invention, humanized immunoglobulins (or antibodies) specific for the alpha-4 subunit of VLA-4 are provided. The humanized immunoglobulins have variable framework regions substantially from a human immunoglobulin (termed an acceptor immunoglobulin) and complementarity determining regions substantially from a mouse immunoglobulin termed mu MAb 21.6 (referred to as the donor immunoglobulin). The constant region(s), if present, are also substantially from a human immunoglobulin. The humanized antibodies exhibit a specific binding affinity for VLA-4 of at least 10^7, 10^8, 10^9, or 10^{10} M^{-1}. Usually the upper limit of binding affinity of the humanized antibodies for VLA-4 is within a factor of three or five of that of mu MAb 21.6 (about 10^9 M^{-1}). Often the lower limit of binding affinity is also within a factor of three or five of that of mu MAb 21.6.

A. General Characteristics of Immunoglobulins

The basic antibody structural unit is known to comprise a tetramer. Each tetramer is composed of two identical pairs of polypeptide chains, each pair having one "light" (about 25 kDa) and one "heavy" chain (about 50-70 kDa). The amino-terminal portion of each chain includes a variable region

of about 100 to 110 or more amino acids primarily responsible for antigen recognition. The carboxy-terminal portion of each chain defines a constant region primarily responsible for effector function.

Light chains are classified as either kappa or lambda. Heavy chains are classified as gamma, mu, alpha, delta, or epsilon, and define the antibody's isotype as IgG, IgM, IgA, IgD and IgE, respectively. Within light and heavy chains, the variable and constant regions are joined by a "J" region of about 12 or more amino acids, with the heavy chain also including a "D" region of about 10 more amino acids. (*See generally, Fundamental Immunology* (Paul, W., ed., 2nd ed. Raven Press, N.Y., 1989), Ch. 7 (incorporated by reference in its entirety for all purposes).

The variable regions of each light/heavy chain pair form the antibody binding site. The chains all exhibit the same general structure of relatively conserved framework regions (FR) joined by three hypervariable regions, also called complementarity determining regions or CDRs. The CDRs from the two chains of each pair are aligned by the framework regions, enabling binding to a specific epitope. CDR and FR residues are delineated according to the standard sequence definition of Kabat et al., *supra*. An alternative structural definition has been proposed by Chothia et al., *J. Mol. Biol.* 196:901-917 (1987); *Nature* 342:878-883 (1989); and *J. Mol. Biol.* 186:651-663 (1989) (hereinafter collectively referred to as "Chothia et al." and incorporated by reference in their entirety for all purposes). When framework positions, as defined by Kabat et al., *supra*, that constitute structural loop positions as defined by Chothia et al., *supra*, the amino acids present in the mouse antibody are usually incorporated into the humanized antibody.

B. Production of Humanized Antibodies

(1) Mouse MAb 21.6

The starting material for production of humanized antibodies is mu MAb 21.6. The isolation and properties of this antibody are described in USSN 07/871,223. Briefly, mu

MAb 21.6 is specific for the alpha-4 subunit of VLA-4 and has been shown to inhibit human lymphocyte binding to tissue cultures of rat brain cells stimulated with tumor necrosis factor. The cloning and sequencing of cDNA encoding the mu MAb 21.6 antibody heavy and light chain variable regions is described in Example 1, and the nucleotide and predicted amino acids sequences are shown in Figures 1 and 2. These figures also illustrate the subdivision of the amino acid coding sequencing into framework and complementarity determining domains. From N-terminal to C-terminal, both light and heavy chains comprise the domains FR1, CDR1, FR2, CDR2, FR3, CDR3 and FR4. The assignment of amino acids to each domain is in accordance with the numbering convention of Kabat et al., *supra*.

(2) Selection of Human Antibodies to Supply Framework Residues

The substitution of mouse CDRs into a human variable domain framework is most likely to result in retention of their correct spatial orientation if the human variable domain framework adopts the same or similar conformation to the mouse variable framework from which the CDRs originated. This is achieved by obtaining the human variable domains from human antibodies whose framework sequences exhibit a high degree of sequence identity with the murine variable framework domains from which the CDRs were derived. The heavy and light chain variable framework regions can be derived from the same or different human antibody sequences. The human antibody sequences can be the sequences of naturally occurring human antibodies or can be consensus sequences of several human antibodies. *See* Kettleborough et al., *Protein Engineering* 4:773 (1991); Kolbinger et al., *Protein Engineering* 6:971 (1993).

Suitable human antibody sequences are identified by computer comparisons of the amino acid sequences of the mouse variable regions with the sequences of known human antibodies. The comparison is performed separately for heavy and light chains but the principles are similar for each. This

comparison reveals that the mu 21.6 light chain shows greatest sequence identity to human light chains of subtype kappa 1, and that the mu 21.6 heavy chain shows greatest sequence identity to human heavy chains of subtype one, as defined by Kabat et al., *supra*. Thus, light and heavy human framework regions are usually derived from human antibodies of these subtypes, or from consensus sequences of such subtypes. The preferred light and heavy chain human variable regions showing greatest sequence identity to the corresponding regions from mu MAb 21.6 are from antibodies RE1 and 21/28'CL respectively.

(3) Computer Modelling

The unnatural juxtaposition of murine CDR regions with human variable framework region can result in unnatural conformational restraints, which, unless corrected by substitution of certain amino acid residues, lead to loss of binding affinity. The selection of amino acid residues for substitution is determined, in part, by computer modelling. Computer hardware and software for producing three-dimensional images of immunoglobulin molecules are widely available. In general, molecular models are produced starting from solved structures for immunoglobulin chains or domains thereof. The chains to be modelled are compared for amino acid sequence similarity with chains or domains of solved three dimensional structures, and the chains or domains showing the greatest sequence similarity is/are selected as starting points for construction of the molecular model. For example, for the light chain of mu MAb 21.6, the starting point for modelling the framework regions, CDR1 and CDR2 regions, was the human light chain RE1. For the CDR3 region, the starting point was the CDR3 region from the light chain of a different human antibody HyHEL-5. The solved starting structures are modified to allow for differences between the actual amino acids in the immunoglobulin chains or domains being modelled, and those in the starting structure. The modified structures are then assembled into a composite immunoglobulin. Finally, the model is refined by energy minimization and by verifying that all atoms are within appropriate distances from one another and

that bond lengths and angles are within chemically acceptable limits. Example 4 discusses in more detail the steps taken to produce a three dimensional computer model for the variable regions of the mu MAb 21.6, and the model is shown in Figure 5. This model can in turn serve as a starting point for predicting the three-dimensional structure of an antibody containing the mu MAb 21.6 complementarity determining regions substituted in human framework structures. Additional models can be constructed representing the structure when further amino acid substitutions to be discussed *infra*, are introduced.

(4) Substitution of Amino Acid Residues

As noted *supra*, the humanized antibodies of the invention comprise variable framework regions substantially from a human immunoglobulin and complementarity determining regions substantially from a mouse immunoglobulin termed mu MAb 21.6. Having identified the complementarity determining regions of mu MAb 21.6 and appropriate human acceptor immunoglobulins, the next step is to determine which, if any, residues from these components should be substituted to optimize the properties of the resulting humanized antibody. In general, substitution of human amino acid residues with murine should be minimized, because introduction of murine residues increases the risk of the antibody eliciting a HAMA response in humans. Amino acids are selected for substitution based on their possible influence on CDR conformation and/or binding to antigen. Investigation of such possible influences is by modelling, examination of the characteristics of the amino acids at particular locations, or empirical observation of the effects of substitution or mutagenesis of particular amino acids.

When an amino acid differs between a mu MAb 21.6 variable framework region and an equivalent human variable framework region, the human framework amino acid should usually be substituted by the equivalent mouse amino acid if it is reasonably expected that the amino acid:

(1) noncovalently binds antigen directly (*e.g.*, amino acids at positions L49, L69 of mu MAb 21.6),

(2) is adjacent to a CDR region, is part of a CDR region under the alternative definition proposed by Chothia et al., *supra*, or otherwise interacts with a CDR region (*e.g.*, is within about 3Å of a CDR region) (*e.g.*, amino acids at positions L45, L58, H27, H28, H29, H30 and H71 of mu MAb 21.6), or

(3) participates in the V_L-V_H interface (*e.g.*, amino acids at position H44 of mu MAb 21.6).

Other candidates for substitution are acceptor human framework amino acids that are unusual for a human immunoglobulin at that position (*e.g.*, amino acids at positions L104, L105 and L107 of mu MAb 21.6). These amino acids can be substituted with amino acids from the equivalent position of more typical human immunoglobulins. Alternatively, amino acids from equivalent positions in the mouse MAb 21.6 can be introduced into the human framework regions when such amino acids are typical of human immunoglobulin at the equivalent positions.

In general, substitution of all or most of the amino acids fulfilling the above criteria is desirable. Occasionally, however, there is some ambiguity about whether a particular amino acid meets the above criteria, and alternative variant immunoglobulins are produced, one of which has that particular substitution, the other of which does not. The humanized antibodies of the present invention will usually contain a substitution of a human light chain framework residue with a corresponding mu MAb 21.6 residue in at least 1, 2 or 3, and more usually 4, of the following positions: L45, L49, L58 and L69. The humanized antibodies also usually contain a substitution of a human heavy chain framework residue in at least 1, 2, 3, 4, or 5, and sometimes 6, of the following positions: H27, H28, H29, H30, H44 and H71. Optionally, H36 may also be substituted. In preferred embodiments when the human light chain acceptor immunoglobulin is RE1, the light chain also contains substitutions in at least 1 or 2, and more usually 3, of the following positions: L104, L105 and L107. These positions are substituted with the amino acid from the equivalent position of a human immunoglobulin having a more

typical amino acid residues. Appropriate amino acids to substitute are shown in Figures 6 and 7.

Usually the CDR regions in humanized antibodies are substantially identical, and more usually, identical to the corresponding CDR regions in the mu MAb 21.6 antibody. Occasionally, however, it is desirable to change one of the residues in a CDR region. For example, Example 5 identifies an amino acid similarity between the mu MAb 21.6 CDR3 and the VCAM-1 ligand. This observation suggests that the binding affinity of humanized antibodies might be improved by redesigning the heavy chain CDR3 region to resemble VCAM-1 even more closely. Accordingly, one or more amino acids from the CDR3 domain can be substituted with amino acids from the VCAM-1 binding domain. Although not usually desirable, it is sometimes possible to make one or more conservative amino acid substitutions of CDR residues without appreciably affecting the binding affinity of the resulting humanized immunoglobulin.

Other than for the specific amino acid substitutions discussed above, the framework regions of humanized immunoglobulins are usually substantially identical, and more usually, identical to the framework regions of the human antibodies from which they were derived. Of course, many of the amino acids in the framework region make little or no direct contribution to the specificity or affinity of an antibody. Thus, many individual conservative substitutions of framework residues can be tolerated without appreciable change of the specificity or affinity of the resulting humanized immunoglobulin. However, in general, such substitutions are undesirable.

(5) Production of Variable Regions

Having conceptually selected the CDR and framework components of humanized immunoglobulins, a variety of methods are available for producing such immunoglobulins. Because of the degeneracy of the code, a variety of nucleic acid sequences will encode each immunoglobulin amino acid sequence. The desired nucleic acid sequences can be produced by *de novo* solid-phase DNA synthesis or by PCR mutagenesis of an earlier

prepared variant of the desired polynucleotide. Oligonucleotide-mediated mutagenesis is a preferred method for preparing substitution, deletion and insertion variants of target polypeptide DNA. *See* Adelman et al., *DNA* 2:183 (1983). Briefly, the target polypeptide DNA is altered by hybridizing an oligonucleotide encoding the desired mutation to a single-stranded DNA template. After hybridization, a DNA polymerase is used to synthesize an entire second complementary strand of the template that incorporates the oligonucleotide primer, and encodes the selected alteration in the target polypeptide DNA.

(6) Selection of Constant Region

The variable segments of humanized antibodies produced as described *supra* are typically linked to at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin. Human constant region DNA sequences can be isolated in accordance with well-known procedures from a variety of human cells, but preferably immortalized B-cells (*see* Kabat et al., *supra,* and WO87/02671) (each of which is incorporated by reference in its entirety for all purposes). Ordinarily, the antibody will contain both light chain and heavy chain constant regions. The heavy chain constant region usually includes CH1, hinge, CH2, CH3, and CH4 regions.

The humanized antibodies include antibodies having all types of constant regions, including IgM, IgG, IgD, IgA and IgE, and any isotype, including IgG1, IgG2, IgG3 and IgG4. When it is desired that the humanized antibody exhibit cytotoxic activity, the constant domain is usually a complement-fixing constant domain and the class is typically IgG_1. When such cytotoxic activity is not desirable, the constant domain may be of the IgG_2 class. The humanized antibody may comprise sequences from more than one class or isotype.

(7) Expression Systems

Nucleic acids encoding humanized light and heavy chain variable regions, optionally linked to constant regions,

are inserted into expression vectors. The light and heavy chains can be cloned in the same or different expression vectors. The DNA segments encoding immunoglobulin chains are operably linked to control sequences in the expression vector(s) that ensure the expression of immunoglobulin polypeptides. Such control sequences include a signal sequence, a promoter, an enhancer, and a transcription termination sequence. Expression vectors are typically replicable in the host organisms either as episomes or as an integral part of the host chromosomal DNA. Commonly, expression vectors will contain selection markers, *e.g.*, tetracycline or neomycin, to permit detection of those cells transformed with the desired DNA sequences (*see, e.g.*, U.S. Patent 4,704,362.)

E. coli is one prokaryotic host useful particularly for cloning the polynucleotides of the present invention. Other microbial hosts suitable for use include bacilli, such as *Bacillus subtilus*, and other enterobacteriaceae, such as *Salmonella, Serratia*, and various *Pseudomonas* species. In these prokaryotic hosts, one can also make expression vectors, which will typically contain expression control sequences compatible with the host cell (*e.g.*, an origin of replication). In addition, any number of a variety of well-known promoters will be present, such as the lactose promoter system, a tryptophan (trp) promoter system, a beta-lactamase promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence, and have ribosome binding site sequences and the like, for initiating and completing transcription and translation.

Other microbes, such as yeast, may also be used for expression. *Saccharomyces* is a preferred host, with suitable vectors having expression control sequences, such as promoters, including 3-phosphoglycerate kinase or other glycolytic enzymes, and an origin of replication, termination sequences and the like as desired.

In addition to microorganisms, mammalian tissue cell culture may also be used to express and produce the

polypeptides of the present invention (*see* Winnacker, *From Genes to Clones* (VCH Publishers, N.Y., N.Y., 1987). Eukaryotic cells are actually preferred, because a number of suitable host cell lines capable of secreting intact immunoglobulins have been developed in the art, and include the CHO cell lines, various Cos cell lines, HeLa cells, preferably myeloma cell lines, or transformed B-cells or hybridomas. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter, and an enhancer (Queen et al., *Immunol. Rev.* 89:49-68 (1986)), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites, and transcriptional terminator sequences. Preferred expression control sequences are promoters derived from immunoglobulin genes, SV40, adenovirus, bovine papilloma virus, cytomegalovirus and the like.

The vectors containing the polynucleotide sequences of interest (*e.g.*, the heavy and light chain encoding sequences and expression control sequences) can be transferred into the host cell by well-known methods, which vary depending on the type of cellular host. For example, calcium chloride transfection is commonly utilized for prokaryotic cells, whereas calcium phosphate treatment or electroporation may be used for other cellular hosts. (*See generally* Sambrook et al., *Molecular Cloning: A Laboratory Manual* (Cold Spring Harbor Press, 2nd ed., 1989) (incorporated by reference in its entirety for all purposes). When heavy and light chains are cloned on separate expression vectors, the vectors are co-transfected to obtain expression and assembly of intact immunoglobulins.

Once expressed, the whole antibodies, their dimers, individual light and heavy chains, or other immunoglobulin forms of the present invention can be purified according to standard procedures of the art, including ammonium sulfate precipitation, affinity columns, column chromatography, gel electrophoresis and the like (*see generally* Scopes, *Protein Purification* (Springer-Verlag, N.Y., 1982). Substantially pure immunoglobulins of at least about 90 to 95% homogeneity are

preferred, and 98 to 99% or more homogeneity most preferred, for pharmaceutical uses.

C. Fragments of Humanized Antibodies

In another embodiment of the invention, fragments of humanized antibodies are provided. Typically, these fragments exhibit specific binding to the VLA-4 antigen with an affinity of at least 10^7 M^{-1}, and more typically 10^8 or 10^9 M^{-1}. Humanized antibody fragments include separate heavy chains, light chains Fab, Fab' $F(ab')_2$, Fabc, and Fv. Fragments are produced by recombinant DNA techniques, or by enzymic or chemical separation of intact immunoglobulins.

II. Nucleic Acids

The humanized antibodies and fragments thereof are usually produced by expression of nucleic acids. All nucleic acids encoding a humanized antibody or a fragment thereof described in this application are expressly included in the invention.

III. Computers

In another aspect of the invention, computers programmed to display three dimensional images of antibodies on a monitor are provided. For example, a Silicon Graphics IRIS 4D workstation running under the UNIX operating system and using the molecular modelling package QUANTA (Polygen Corp. USA) is suitable. Computers are useful for visualizing models of variants of humanized antibodies. In general, the antibodies of the invention already provide satisfactory binding affinity. However, it is likely that antibodies with even stronger binding affinity could be identified by further variation of certain amino acid residues. The three dimensional image will also identify many noncritical amino acids, which could be the subject of conservative substitutions without appreciable affecting the binding affinity of the antibody. Collectively even conservative substitutions can have a significant effect on the properties of an immunoglobulin. However, it is likely many individual

conservative substitutions will not significantly impair the properties of the immunoglobulins.

IV. Testing Humanized Antibodies

The humanized antibodies of the invention are tested by a variety of assays. These include a simple binding assay for detecting the existence or strength of binding of an antibody to cells bearing the VLA-receptor. The antibodies are also tested for their capacity to block the interaction of cells bearing the VLA-4 receptor with endothelial cells expressing a VCAM-1 ligand. The endothelial cells may be grown and stimulated in culture or may be a component of naturally occurring brain tissue sections. *See* Yednock et al., *supra,* and USSN 07/871,223. The humanized antibodies are also tested for their capacity to prevent or reduce inflammation and subsequent paralysis in laboratory animals having experimental autoimmune encephalomyelitis (EAE). EAE is induced by injection of a laboratory animal with $CD4^+$ T-cells specific for myelin basic protein or by directly immunizing animals with myelin basic protein. This protein is localized in the central nervous system, and the reactive T-cells initiate destruction of sheaths containing this protein in a manner that simulates the autoimmune response in multiple sclerosis. *See* Yednock et al., *supra,* and copending USSN 07/871,223.

V. Pharmaceutical Compositions

The invention provides pharmaceutical compositions to be used for prophylactic or therapeutic treatment comprising an active therapeutic agent, *i.e.,* a humanized 21.6 antibody or a binding fragment thereof, and a variety of other components. The preferred form depends on the intended mode of administration and therapeutic application. The compositions may also include, depending on the formulation desired, pharmaceutically-acceptable, non-toxic carriers or diluents, which are defined as vehicles commonly used to formulate pharmaceutical compositions for animal or human administration. The diluent is selected so as not to affect the biological activity of the combination. Examples of such diluents are

distilled water, physiological phosphate-buffered saline, Ringer's solutions, dextrose solution, and Hank's solution. In addition, the pharmaceutical composition or formulation may also include other carriers, adjuvants, or nontoxic, nontherapeutic, nonimmunogenic stabilizers and the like.

VI. Methods of Diagnosis

The humanized antibodies and their binding fragments are useful for detecting the presence of cells bearing the VLA-4 receptor. The presence of such cells in the brain is diagnostic of an inflammatory response and may signal the need for commencement of a therapeutic method discussed *infra*. Diagnosis can be accomplished by removing a cellular sample from a patient. The amount of expressed VLA-4 antigen in individual cells of the sample is then determined, *e.g.*, by immunohistochemical staining of fixed cells or by Western blotting of a cell extract with a humanized MAb 21.6 antibody or a binding fragment thereof.

Diagnosis can also be achieved by *in vivo* administration of a labelled humanized MAb 21.6 (or binding fragment) and detection by *in vivo* imaging. The concentration of humanized MAb 21.6 administered should be sufficient that the binding to cells having the target antigen is detectable compared to the background signal. The diagnostic reagent can be labelled with a radioisotope for camera imaging, or a paramagnetic isotope for magnetic resonance or electron spin resonance imaging.

A change (typically an increase) in the level of VLA-4 protein in a cellular sample or imaged from an individual, which is outside the range of clinically established normal levels, may indicate the presence of an undesirable inflammatory response reaction in the individual from whom the sample was obtained, and/or indicate a predisposition of the individual for developing (or progressing through) such a reaction. VLA-4 protein can also be employed as a differentiation marker to identify and type cells of certain lineages and developmental origins. Such cell-type specific

detection can be used for histopathological diagnosis of undesired immune responses.

VII. Methods of Treatment

The invention also provides methods of treatment that exploit the capacity of humanized MAb 21.6 to block α4-dependent interactions of the VLA-4 receptor. The α4-dependent interaction of the VLA-4 receptor with the VCAM-1 ligand on endothelial cells is an early event in many inflammatory responses, particularly those of the central nervous system. Undesired diseases and conditions resulting from inflammation of the central nervous system include acute diseases, such as stroke and other cerebral traumas, and chronic diseases, such as multiple sclerosis, meningitis and encephalitis. Multiple sclerosis is a progressive neurological autoimmune disease that affects an estimated 250,000 to 350,000 people in the United States. Multiple sclerosis is thought to be a the result of a specific autoimmune reaction in which certain leukocytes attack and initiate the destruction of myelin, the insulating sheath covering nerve fibers. In an animal model for multiple sclerosis, murine monoclonal antibodies directed against alpha-4-beta-1 integrin have been shown to block the adhesion of leukocytes to the endothelium, and thus prevent inflammation of the central nervous system and subsequent paralysis in the animals.

The humanized MAb 21.6 antibodies of the present invention offer several advantages over the mouse antibodies already shown to be effective in animals models:

1) The human immune system should not recognize the framework or constant region of the humanized antibody as foreign, and therefore the antibody response against such an injected antibody should be less than against a totally foreign mouse antibody or a partially foreign chimeric antibody.

2) Because the effector portion of the humanized antibody is human, it may interact better with other parts of the human immune system.

3) Injected mouse antibodies have been reported to have a half-life in the human circulation much shorter than the

half-life of normal human antibodies (Shaw et al., *J. Immunol.* 138:4534-4538 (1987)). Injected humanized antibodies have a half-life essentially equivalent to naturally occurring human antibodies, allowing smaller and less frequent doses.

The pharmaceutical compositions discussed *supra* can be administered for prophylactic and/or therapeutic treatments of multiple sclerosis or other inflammatory disorders, particularly those of the central nervous system. In therapeutic applications, compositions are administered to a patient suspected of, or already suffering from a disease such as multiple sclerosis, in an amount sufficient to cure, or at least partially arrest, the symptoms of the disease and its complications. An amount adequate to accomplish this is defined as a therapeutically- or pharmaceutically-effective dose.

In prophylactic applications, pharmaceutical compositions are administered to a patient susceptible to, or otherwise at risk of, a particular disease in an amount sufficient to eliminate or reduce the risk or delay the outset of the disease. Such an amount is defined to be a prophylactically effective dose. In patients with multiple sclerosis in remission, risk may be assessed by NMR imaging or, in some cases, by presymptomatic indications observed by the patient.

The pharmaceutical compositions will be administered by parenteral, topical, intravenous, oral, or subcutaneous, intramuscular local administration, such as by aerosol or transdermally, for prophylactic and/or therapeutic treatment. The pharmaceutical compositions can be administered in a variety of unit dosage forms depending upon the method of administration. For example, unit dosage forms suitable for oral administration include powder, tablets, pills, capsules, and lozenges.

Effective doses of the compositions of the present invention, for the treatment of the above described conditions will vary depending upon many different factors, including means of administration, target site, physiological state of the patient, and other medicants administered. Thus, treatment

dosages will need to be titrated to optimize safety and efficacy. These compositions may be administered to mammals for veterinary use and for clinical use in humans in a manner similar to other therapeutic agents, *i.e.*, in a physiologically acceptable carrier. In general, the administration dosage will range from about 0.0001 to 100 mg/kg, and more usually 0.01 to 0.5 mg/kg of the host body weight.

VIII. Other Uses

The humanized antibodies are also useful for affinity purification of the VLA-4 receptor. The antibodies are immobilized to a solid support and a solution of dispersed proteins is passed over the support. VLA-4 binds to the support and is thereby separated from other proteins. The purified VLA-4 or a fragment thereof, made available by this method, can be used as a vaccine or as an immunogen for producing further antibodies.

The humanized antibodies of the invention are also useful for generating idiotypic antibodies by, for example, immunization of an animal with a humanized antibody. An anti-idiotype antibody whose binding to the human antibody is inhibited by VLA-4 or fragments thereof is selected. Because both the anti-idiotypic antibody and the VLA-4 or fragments thereof bind to the humanized immunoglobulin, the anti-idiotypic antibody may represent the "internal image" of an epitope and thus may substitute the ligand of the VLA-4 receptor, i.e., VCAM-1.

EXAMPLES

Example 1: Cloning and Sequencing of the Mouse 21.6 Variable Regions

The mouse anti-VLA antibody 21.6 has been described in co-pending application USSN 07/871,223. Total RNA was isolated from hybridoma cells producing mouse 21.6 antibody. First-strand cDNA was synthesized using a kit (Pharmacia Biosystems Limited). Heavy and light chain variable regions were obtained by using PCR primers designed to hybridize to sequences flanking and external to the sequences coding for the

variable regions, thereby allowing cloning of the entire coding sequences for the mouse 21.6 antibody variable regions. Sense PCR primers hybridizing to the 5'-ends of mouse kappa light-chain leader sequences and of mouse heavy-chain leader sequences were designed based on databases of 42 mouse kappa light-chain leader sequences and of 55 mouse heavy-chain leader sequences (Jones & Bendig, *Bio/Technology* 9:88-89 (1991) (incorporated by reference in its entirety for all purposes)). These primers were used in conjunction with anti-sense PCR primers hybridizing to the 3'-ends of the mouse constant regions (kappa or gamma).

Mouse 21.6 kappa V_L regions were PCR-amplified in a 50 μl reaction typically containing 10 mM Tris-HCl (pH 8.3), 50 mM KCI, 200 μM dNTPs, 1.5 mM $MgCl_2$, 1 unit of AmpliTaq (Perkin Elmer Cetus) DNA polymerase, 1 μl of cDNA template, 0.25 μM of MKV primer and 0.25 μM of mouse kappa light chain anti-sense PCR primer (Figure 1). Mouse 21.6 V_H regions were PCR-amplified as described above except that MHVH primer and an anti-sense PCR primer specific for the mouse IgG1 heavy chain constant region were used (Figure 2). Each PCR reaction was cycled, after an initial melt at 94°C for 5 min, at 94°C for 1 min, 55°C for 1 min, and 72°C for 2 min over 25 cycles. The completion of the last cycle was followed by a final extension at 72°C for 10 min. The ramp time between the primer-annealing and extension steps was 2.5 min. Following PCR amplification, 10 μl aliquots from each reaction were analyzed on ethidium-bromide-stained 1.5% agarose gels.

The PCR products were cloned using the "TA Cloning System" (Invitrogen Corporation). Vectors containing inserts of the correct size were sequenced using double-stranded plasmid DNA and Sequenase (United States Biochemical Corporation). To avoid any errors that might have been introduced during the PCR amplification steps, at least two independently PCR-amplified and cloned DNA fragments were sequenced for each variable region.

The sequences of PCR products were compared with other mouse light chain and heavy chain variable regions (*see* Tables 1 and 2). This comparison indicated that the PCR

products from MKV2 and MKV4 primers represent authentic mouse 21.6 kappa variable regions, and those from MHV1 and MHV2 primers represent authentic mouse V_H regions, and it was concluded that the sequences of these product are those of the mouse 21.6 antibody variable regions. The DNA and amino acid sequences of the cDNA coding for the mouse 21.6 V_L and V_H regions are shown in Figures 1 and 2.

Table 1

Comparison of the mouse 21.6 light chain variable region to other light chain variable regions.

Mouse 21.6 V_L versus:	Percent Similarity[1]	Percent Identity
Consensus sequence for mouse kappa V_L subgroup 5[2]	84.0	72.6
Consensus sequence for human kappa V_L subgroup 1[2]	84.0	69.8
Consensus sequence for human kappa V_L subgroup 2[2]	65.1	52.8
Consensus sequence for human kappa V_L subgroup 3[2]	72.6	57.5
Consensus sequence for human kappa V_L subgroup 4[2]	72.6	58.5
Sequence of V_L from human REI[3] (Member of human kappa V_L subgroup 1)	81.0	72.4

[1]Percent similarity was determined using the "GAP" program of the University of Wisconsin Genetics Computer Group.

[2]Consensus sequences were taken from Kabat et al., *supra*.

[3]REI as sequenced by Palm et al., *Hoppe-Seyler's Z. Physiol. Chem.* 356:167-191 (1975).

Table 2

Comparison of the mouse 21.6 heavy chain variable region to other heavy chain variable regions.

Mouse 21.6 V_H versus:	Percent Similarity[1]	Percent Identity
Consensus sequence for mouse V_H subgroup 2c[2]	94.3	91.1
Consensus sequence for human V_H subgroup 1[2]	78.0	65.0
Consensus sequence for human V_H subgroup 2[2]	70.5	53.3
Consensus sequence for human V_H subgroup 3[2]	67.5	52.8
Sequence of V_H from human 21/28'CL[3] (Member of human V_H subgroup 1)	76.5	64.7

[1]Percent similarity was determined using the "GAP" program of the University of Wisconsin Genetics Computer Group.

[2]Consensus sequences were taken from Kabat et al., *supra*.

[3]21/28'CL as sequenced by Dersimonian et al., *J. Immunol.* 139:2496-2501 (1987).

Example 2: Construction of Chimeric 21.6 Antibody

Chimeric light and heavy chains were constructed by linking the PCR-cloned cDNAs of mouse 21.6 V_L and V_H regions to human constant regions. The 5'- and 3'-ends of the mouse cDNA sequences were modified using specially designed PCR primers. The 5'-end PCR-primers (Table 3), which hybridize to the DNA sequences coding for the beginnings of the leader sequences, were designed to create the DNA sequences essential for efficient translation (Kozak, *J. Mol. Biol.* 196:947-950 (1987)), and to create a HindIII restriction sites for cloning into an expression vector. The 3'-end primers (Table 3), which hybridize to the DNA sequences coding for the ends of J

regions, were designed to create the DNA sequences essential for splicing to the constant regions, and to create a BamHI site for cloning into an expression vector. The products of PCR amplification were digested with HindIII and BamHI, cloned into a pUC19 vector, and sequenced to confirm that no errors had occurred during PCR amplification. The adapted mouse 21.6 variable regions were then subcloned into mammalian cells expression vectors containing either the human kappa or gamma-1 constant regions (Figure 3).

Table 3

PCR primers for the construction of chimeric 21.6 antibody.

A. Light chain variable region

1. Primer for reconstruction of the 5'-end (37mer) (SEQ ID NO: 18)

5' C AGA AAG CTT **GCC GCC ACC** ATG AGA CCG TCT ATT CAG 3'
HindIII Kozak Consensus Sequence M R P S I Q

2. Primer for reconstruction of the 3'-end (35mer) (SEQ ID NO: 19)

5' CC GAG GAT CCA **CTC ACG** TTT GAT TTC CAG CTT GGT 3'
BamHI Splice donor site

B. Heavy chain variable region

1. Primer for reconstruction of the 5'-end (37mer) (SEQ ID NO: 20)

5' C AGA AAG CTT **GCC GCC ACC** ATG AAA TGC AGC TGG GTC 3'
HindIII Kozak Consensus Sequence M K C S W V

2. Primer for reconstruction of the 3'-end (33mer) (SEQ ID NO: 21)

5' CC GAG GAT CCA **CTC ACC TGA** GGA GAC GGT GAC T 3'
BamHI Splice donor site

Example 3: Expression and Analysis of 21.6 Chimeric Antibody

The two plasmid DNAs coding for the chimeric 21.6 light and heavy chains were cotransfected into Cos cells. After two or three days, media from the Cos cells was analyzed

by ELISA (1) for the production of a human IgG-like antibody and (2) for the ability of this human-like antibody to bind to L cells expressing human $\alpha 4\beta 1$ integrin on their surface. Figures 4 and 12 show analyses of unpurified and protein-A purified samples of chimeric 21.6 antibody for binding to human $\alpha 4\beta 1$ integrin, in comparison with purified mouse 21.6 antibody control. These figures show that the chimeric 21.6 antibody bound well to antigen and confirm that the correct mouse 21.6 V_L and V_H regions had been cloned.

Example 4: Modelling the Structure of the Mouse 21.6 Variable Regions

A molecular model of the V_L and V_H regions of mouse 21.6 antibody was built. The model was built on a Silicon Graphics IRIS 4D workstation running under the UNIX operating system and using the molecular modelling package QUANTA (Polygen Corp., USA). The structure of the FRs of mouse 21.6 V_L region was based on the solved structure of human Bence-Jones immunoglobulin REI (Epp et al., *Biochemistry* 14:4943-4952 (1975)). The structure of the FRs of mouse 21.6 V_H region was based on the solved structure of mouse antibody Gloop2. Identical residues in the FRs were retained; non-identical residues were substituted using the facilities within QUANTA. CDR1 and CDR2 of mouse 21.6 V_L region were identified as belonging to canonical structure groups 2 and 1, respectively (Chothia et al., *supra)*. Since CDR1 and CDR2 of REI belong to the same canonical groups, CDR1 and CDR2 of mouse 21.6, V_L region were modelled on the structures of CDR1 and CDR2 of REI. CDR3 of mouse 21.6 V_L region did not appear to correspond to any of the canonical structure groups for CDR3s of V_L regions. A database search revealed, however, that CDR3 in mouse 21.6 V_L region was similar to CDR3 in mouse HyHEL-5 V_L region (Sheriff et al., *Proc. Natl. Acad. Sci. USA* 84:8075-8079 (1987)). Thus, the CDR3 of mouse 21.6 V_L region was modelled on the structure of CDR3 in mouse HyHEL-5 V_L region. CDR1 and CDR2 of mouse 21.6 V_H region were identified as belonging to canonical structure groups 1 and 2, respectively. CDR1 of mouse 21.6 V_H region was modelled on CDR1 of Gloop2 V_H region which closely

resembles members of canonical group 1 for CDR1s of V_H regions. CDR2 of mouse 21.6 V_H region was modelled on CDR2 of mouse HyHEL-5 (Sheriff et al., *supra*), which is also a member of canonical group 2 for CDR2 for V_H regions. For CDR3s of V_H regions, there are no canonical structures. However, CDR3 in mouse 21.6 V_H region was similar to CDR3 in mouse R19.9 V_H region (Lascombe et al., *Proc. Natl. Acad. Sci. USA* 86:607-611 (1989)) and was modelled on this CDR3 by by removing an extra serine residue present at the apex of the CDR3 loop of mouse R19.9 V_H region and annealing and refining the gap. The model was finally subjected to steepest descents and conjugate gradients energy minimization using the CHARMM potential (Brooks et al., *J. Comp. Chem.* 4:187-217 (1983)) as implemented in QUANTA in order to relieve unfavorable atomic contacts and to optimize van der Waals and electrostatic interactions.

A view of the structural model of the mouse 21.6 variable regions is presented in Figure 5. The model was used to assist in refining the design of the humanized 21.6 antibody variable regions.

Example 5: Design of Reshaped Human 21.6 Variable Regions

(1) Selection of Homologous Human Antibodies for Framework Sequence

Human variable regions whose FRs showed a high percent identity to those of mouse 21.6 were identified by comparison of amino acid sequences. Tables 4 and 5 compare the mouse 21.6 variable regions to all known mouse variable regions and then to all known human variable regions. The mouse 21.6 V_L region was identified as belonging to mouse kappa V_L region subgroup 5 as defined by Kabat et al., *supra*. Individual mouse kappa V_L regions were identified that had as much as 93.4% identity to the mouse 21.6 kappa V_L region (38C13V'CL and PC613'CL). Mouse 21.6 V_L region was most similar to human kappa V_L regions of subgroup 1 as defined by Kabat et al., *supra*. Individual human kappa V_L regions were identified that had as much as 72.4% identity to the mouse 21.6 kappa V_L region. The framework regions (FRs) from one of the most similar human variable regions, REI, were used in the design of

reshaped human 21.6 V_L region. Mouse 21.6 V_H region was identified as belonging to mouse V_H region subgroup 2c as defined by Kabat et al., *supra*. Individual mouse heavy chain variable regions were identified that have as much as 93.3 % identity to the mouse 21.6 V_H region (17.2.25'CL and 87.92.6'CL). Mouse 21.6 V_H region was most similar to human V_H regions of subgroup 1 as defined by Kabat et al., *supra*. Individual human V_H regions were identified that had as much as 64.7% identity to the mouse 21.6 V_H region. The FRs from one of the most similar human variable regions, 21/28'CL, was used in the design of reshaped human 21.6 V_H region.

(2) Substitution of Amino Acids in Framework Regions

(a) Light Chain

The next step in the design process for the reshaped human 21.6 V_L region was to join the CDRs from mouse 21.6 V_L region to the FRs from human REI (Palm et al., *supra*). In the first version of reshaped human 21.6 V_L region (La), seven changes were made in the human FRs (Table 4, Figure 6).

At positions 104, 105, and 107 in FR4, amino acids from RE1 were substituted with more typical human J region amino acids from another human kappa light chain (Riechmann et al., *Nature* 332:323-327 (1988)).

At position 45 in FR2, the lysine normally present in REI was changed to an arginine as found at that position in mouse 21.6 V_L region. The amino acid residue at this position was thought to be important in the supporting the CDR2 loop of the mouse 21.6 V_L region.

At position 49 in FR2, the tyrosine normally present in REI was changed to an histidine as found at that position in mouse 21.6 V_L region. The histidine at this position in mouse 21.6 V_L region was observed in the model to be located in the middle of the binding site and could possibly make direct contact with antigen during antibody-antigen binding.

At position 58 in FR3, the valine normally present in REI was changed to an isoleucine as found at that position in mouse 21.6 V_L region. The amino acid residue at this position

was thought to be important in the supporting the CDR2 loop of the mouse 21.6 V_L region.

At position 69 in FR3, the threonine normally present in REI was changed to an arginine as found at that position in mouse 21.6 V_L region. The arginine at this position in mouse 21.6 V_L region was observed in the model to be located adjacent to the CDR1 loop of mouse 21.6 V_L region and could possibly make direct contact with the antigen during antibody-antigen binding.

A second version of reshaped human 21.6 V_L region (termed Lb) was designed containing the same substitutions as above except that no change was made at position 49 in FR2 of REI. (Figure 6).

(b) Heavy Chain

The next step in the design process for the reshaped human 21.6 V_H region was to join the CDRs from mouse 21.6 V_H region to the FRs from 21/28'CL (Dersimonian et al., *J. Immunol.* 139:2496-2501 (1987)). In the first version of reshaped human 21.6 V. region (Ha), five changes were made in the human framework regions (Table 5, Figure 7). The five changes in the human FRs were at positions 27, 28, 29, 30, and 71.

At positions 27, 28, 29, and 30 in FR1, the amino acids present in human 21/28'CL were changed to the amino acids found at those positions in mouse 21.6 V_H region. Although these positions are designated as being within FR1 (Kabat et al., *supra*), positions 26 to 30 are part of the structural loop that forms the CDR1 loop of the V_H region. It is likely, therefore, that the amino acids at these positions are directly involved in binding to antigen. Indeed, positions 27 to 30 are part of the canonical structure for CDR1 of the V_H region as defined by Chothia et al., *supra*.

At position 71 in FR3, the arginine present in human 21/28'CL was changed to a alanine as found at that position in mouse 21.6 V_H region. Position 71 is part of the canonical structure for CDR2 of the V_H region as defined by Chothia et al., *supra*. From the model of the mouse 21.6 variable regions,

it appears that the alanine at position 71 is important in supporting the CDR2 loop of the V_H region. A substitution of an arginine for an alanine at this position would very probably disrupt the placing of the CDR2 loop.

A second version (Hb) of reshaped human 21.6 V_H region contains the five changes described above for version Ha were made plus one additional change in FR2.

At position 44 in FR2, the arginine present in human 21/28'CL was changed to a glycine as found at that position in mouse 21.6 V_H region. Based on published information on the packing Of V_L-V_H regions and on the model of the mouse 21.6 variable regions, it was thought that the amino acid residue at position 44 might be important in the packing of the V_L-V_H regions (Chothia et al., *supra*) (Figure 5).

Reshaped human 21.6 V. region version Hc was designed to make the CDR3 loop look more similar to human VCAM-1. Both mouse 21.6 antibody and human VCAM-1 bind to the $\alpha 4\beta 1$ integrin. The CDR3 loop of the V_H region of antibodies is the most diverse of the six CDR loops and is generally the most important single component of the antibody in antibody-antigen interactions (Chothia et al., *supra*; Hoogenboom & Winter, *J. Mol. Biol.* 227:381-388 (1992); Barbas et al., *Proc. Natl. Acad. Sci. USA* 89:4457-4461 (1992)). Some sequence similarity was identified between the CDR3 of mouse 21.6 V_H region and amino acids 86 to 94 of human VCAM-1, particularly, between the YGN (Tyrosine-Glycine-Asparagine) sequence in the CDR3 loop and the FGN (Phenylalanine-Glycine-Asparagine) sequence in VCAM-1. These sequences are thought to be related to the RGD (Arginine-Glycine-Aspartic acid) sequences important in various cell adhesion events (Main et al., *Cell* 71:671-678 (1992)). Therefore, at position 98 in CDR3, the tyrosine present in mouse 21.6 V_H region was changed to a phenylalanine as found in the sequence of human VCAM-1.

Possible substitution at position 36 in FR2 was also considered. The mouse 21.6 VH chain contains an unusual cysteine residue at position 36 in FR2. This position in FR2 is usually a tryptophan in related mouse and human sequences (Table 5). Although cysteine residues are often important for

conformation of an antibody, the model of the mouse 21.6 variable regions did not indicate that this cysteine residue was involved either directly or indirectly with antigen binding so the tryptophan present in FR2 of human 21/28'CL V_H region was left unsubstituted in all three versions of humanized 21.6 antibody.

Example 6: Construction of Reshaped Human 21.6 Antibodies

The first version of reshaped human 21.6 V_L region (resh21.6VLa) was constructed from overlapping PCR fragments essentially as described by Daugherty et al., *Nucleic Acids Res.* 19:2471-2476 (1991). (*See* Figure 8). The mouse 21.6 V_L region, adapted as described in Example 2 and inserted into pUC19, was used as a template. Four pairs of primers, APCR1-vla1, vla2-vla3, vla4-vla5, and vla6-vla7 were synthesized (Table 6 and Figure 8). Adjacent pairs overlapped by at least 21 bases. The APCR1 primer is complementary to the pUC19 vector. The appropriate primer pairs (0.2 μmoles) were combined with 10 ng of template DNA, and 1 unit of AmpliTaq DNA polymerase (Perkin Elmer Cetus) in 50 μl of PCR buffer containing 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 200 μM dNTPs, and 1.5 mM $MgCl_2$. Each reaction was carried out for 25 cycles. After an initial melt at 94° for 5 min, the reactions were cycled at 94°C for 1 min, 55°C for 1 min, and 72°C for 2 min, and finally incubated at 72°C for a further 10 min. The ramp time between the primer-annealing and extension steps was 2.5 min. The products of the four reactions (A, B, C, and D) from the first round of PCR reactions were phenol-extracted and ethanol-precipitated.

Table 6
PCR primers for the construction of reshaped human 21.6 variable regions.

A. Light chain variable region

1. Primers for the synthesis of version "a"

21.6VLa1 (39mer): (SEQ ID NO:22):
5' GAT GGT GAC TCT ATC TCC TAC AGA TGC AGA CAG TGA GGA 3'

21.6VLa2 (32mer): (SEQ ID NO:23):
5' CTG TAG GAG ATA GAG TCA CCA TCA CTT GCA AG 3'

21.6VLa3 (39mer): (SEQ ID NO:24):
5' AGG AGC TTT TCC AGG TGT CTG TTG GTA CCA AGC CAT ATA 3'

21.6VLa4 (41mer): (SEQ ID NO:25):
5' ACC AAC AGA CAC CTG GAA AAG CTC CTA GGC TGC TCA TAC AT 3'

21.6VLa5 (40mer): (SEQ ID NO:26):
5' GCA GGC TGC TGA TGG TGA AAG TAT AAT CTC TCC CAG ACC C 3'

21.6VLa6 (42mer): (SEQ ID NO:27):
5' ACT TTC ACC ATC AGC AGC CTG CAG CCT GAA GAT ATT GCA ACT 3'

21.6VLa7 (59mer): (SEQ ID NO:28):
5' CCG AGG ATC CAC TCA CGT TTG ATT TCC ACC TTG GTG CCT TGA CCG AAC GTC CAC AGA TT 3'

2. Primers for the synthesis of version "b"

(SEQ ID NO:29)
21.6VLb1 (33mer): changes H-49 to Y-49
5' GGA AAA GCT CCT AGG CTG CTC ATA TAT TAC ACA 3'

(SEQ ID NO:30)
21.6VLb2 (38mer): changes ACC-101 to ACA-101 to destroy an StyI site
5' CCG AGG ATC CAC TCA CGT TTG ATT TCC ACC TTT GTG CC 3'

B. Heavy chain variable region

1. Primers for the synthesis of version "a"

21.6VHa1 (51mer): (SEQ ID NO:31):
5' AAC CCA GTG TAT ATA GGT GTC TTT AAT GTT GAA ACC GCT AGC TTT ACA GCT 3'

21.6VHa2 (67mer): (SEQ ID NO:32):
5' AAA GAC ACC TAT ATA CAC TGG GTT AGA CAG GCC CCT GGC CAA AGG CTG GAG TGG ATG GGA AGG ATT G 3'

21.6VHa3 (26mer): (SEQ ID NO:33):
5' GAC CCG GCC CTG GAA CTT CGG GTC AT 3'

21.6VHa4 (66mer): (SEQ ID NO:34):
5' GAC CCG AAG TTC CAG GGC CGG GTC ACC ATC ACC GCA GAC ACC TCT GCC AGC ACC GCC TAC ATG GAA 3'

21.6VHa5 (64mer): (SEQ ID NO:35):
5' CCA TAG CAT AGA CCC CGT AGT TAC CAT AAT ATC CCT CTC TGG CGC AGT AGT AGA CTG CAG TGT C 3'

21.6VHa6 (63mer) (SEQ ID NO:36):
5' GGT AAC TAC GGG GTC TAT GCT ATG GAC TAC TGG GGT CAA GGA ACC CTT GTC ACC GTC TCC TCA 3'

2. Primer for the synthesis of version "b"

21.6VHb (37mer) (SEQ ID NO:37): changes R-44 to G-44
5' CCA GGG CCG GGT CAC CAT CAC CAG AGA CAC CTC TGC C 3'

3. Primer for the synthesis of version "c"

21.6VHc (27mer) (SEQ ID NO:38): changes Y-98 to F-98
5' CAG GCC CCT GGC CAA GGG CTG GAG TGG 3'

C. Both light and heavy chain variable regions

Primers hybridizing to the flanking pUC19 vector DNA

APCR1 (17mer, sense primer) (SEQ ID NO:39)
5' TAC GCA AAC CGC CTC TC 3'

APCR4 (18mer, anti-sense primer) (SEQ ID NO:40)
5' GAG TGC ACC ATA TGC GGT 3'

PCR products A and B, and C and D were joined in a second round of PCR reactions. PCR products A and B, and C and D, (50 ng of each) were added to 50 μl PCR reactions (as described above) and amplified through 20 cycles as described above, except that the annealing temperature was raised to 60°C. The products of these reactions were termed E and F. The pairs of PCR primers used were APCR1-vla3 and vla4-vla7, respectively. PCR products E and F were phenol-extracted and ethanol-precipitated and then assembled in a third round of PCR reactions by their own complementarity in a two step-PCR reaction similar to that described above using APCR1 and vla7 as the terminal primers. The fully assembled fragment representing the entire reshaped human 21.6 V_L region including a leader sequence was digested with HindIII and BamHI and cloned into pUC19 for sequencing. A clone having the correct sequence was designated resh21.6VLa.

The second version of a reshaped human 21.6 V_L region (Lb) was constructed using PCR primers to make minor modifications in the first version of reshaped human 21.6 V_L region (La) by the method of Kamman et al., *Nucl. Acids Res.*

17:5404 (1989). Two sets of primers were synthesized (Table 6). Each PCR reaction was essentially carried out under the same conditions as described above. In a first PCR reaction, mutagenic primer 21.6VLb2 was used to destroy a StyI site (Thr-ACC-97 to Thr-ACA-97) to yield resh21.6VLa2. Then, in a second PCR reaction, mutagenic primer 21.6VLb1 (His-49 to Tyr-49) was used with pUC-resh21.6VLa2 as template DNA. The PCR product was cut with StyI and BamHI and subcloned into pUC-resh21.6VLa2, cleaved with the same restriction enzymes. A clone with the correct sequence was designated pUC-resh21.6VLb.

Version "a" of a reshaped human 21.6 V_H region was constructed using the same PCR methods as described for the construction of version "a" of reshaped human 21.6 V_L region (Table 6 and Figure 9). The HindIII-BamHI DNA fragments coding for version "g" of reshaped human 425 V_H region (Kettleborough et al., *supra*) and version "b" of reshaped human AUK12-20 V_H region were subcloned into pUC19 vectors yielding pUC-resh425g and pUC-reshAUK12-20b, respectively. (Version "b" of AUK12-20, was derived by PCR mutagenesis of a fragment V_Ha425 described by Kettleborough et al., *supra*, and encodes the amino acid sequence (SEQ ID NO: 41):

QVQLVQSGAEVKKPGASVKVSCKASGYSFT SYYIH WVRQAPGQGLEWVG

YIDPFNGGTSYNQKFKG KVTMTVDTSTNTAYMELSSLRSEDTAVYYCAR GGN-RFAY WGQGTLVTVSS

(spaces separate FR and CDR regions)).

Plasmid pUC-resh425g annd pUC-reshAUK12-20b, as well as the pUC vector containing the mouse 21.6 V_H region as modified for use in the construction of the chimeric 21.6 heavy chain (pUC-chim21.6VH), were used as template DNAs in the subsequent PCR reactions. PCR primers were designed and synthesized for the construction of version "a" of reshaped human 21.6 V_H region (Table 6). PCR product A (Figure 9) was obtained using pUC-reshAUK12-20b as DNA template and APCR1-vha1 as the PCR primer pair. PCR products B and D were obtained using pUC-chim21.6VH as DNA template and vha2-vha3 and vha6-APCR4 as PCR primer pairs, respectively. Finally, PCR product C was obtained using pUC-resh425g as DNA template and vla4-vla5 as the PCR primer pair. The final PCR product was subcloned into pUC19 as an

HindIII-BamHI fragment for DNA sequencing. A clone with the correct DNA sequence was designated pUC-resh21.6VHa. The DNA and amino acid sequences of the first version of the reshaped 21.6 variable region are shown in Figure 10.

The remaining versions of reshaped human 21.6 V_H region were constructed essentially as described above for the construction of version "b" of reshaped human 21.6 V_L region. Two sets of primers were synthesized (Table 6). For the second (Hb) and third (Hc) versions, mutagenic primers 21.6VHb (Arg-44 to Gly-44) and 21.6VHc (Tyr-98 to Phe-98), respectively, were used in PCR reactions with pUC-resh21.6VHa as the template DNA. The PCR products VHb and VHc were cut with restriction enzymes and subcloned into pUC vector pUC-resh21.6VHa as MscI-BamHI and PstI-BamHI fragments, respectively, to yield pUC-resh21.6VHb and pUC-resh21.6VHc.

The first version of a reshaped human 21.6 V_H region (Ha) was constructed in a similar manner to that used for the construction of the first version of reshaped human 21.6 V_L region (La). In this case, however, PCR primers were used with three different template DNAs, mouse 21.6 V_H region as already adapted for expression of chimeric 21.6 heavy chain, humanized 425 V_H region version "g" (Kettleborough et al., *supra*), and humanized AUK12-20 version "b" V_H region (Table 6, Figure 9). The DNA and amino acid sequences of the first version of the humanized 21.6 heavy chain variable region are shown in Figure 11. The second and third versions of a humanized 21.6 V_H region (Hb and Hc) were constructed using PCR primers to make minor modifications in the first version of humanized 21.6 V_H region (Ha) (Table 6).

Example 7: Expression and Analysis of Humanized Antibodies

1. Linkage of Variable Regions to Constant Regions in Expression Vectors

The DNA fragments coding for the chimeric and reshaped 21.6 V_L and V_H regions were subcloned into HCMV vectors designed to express either human kappa light chains or human gamma-1 heavy chains in mammalian cells (*see* Figure 3) and Maeda et al., *Hum. Antibod. Hybridomas* 2:124-134 (1991).

Both vectors contain the human cytomegalovirus (HCMV) promoter and enhancer for high level transcription of the immunoglobulin light and heavy chains. The light chain expression vector is exactly as described in Maeda et al., *supra,* and contains genomic DNA coding for the human kappa constant region (Rabbitts et al., *Curr. Top. Microbiol. Immunol.* 113:166-171 (1984)). The heavy chain expression vector is essentially as described in Maeda et al., *supra,* with the exception that the genomic DNA coding for the human gamma-1 constant region was replaced with a cDNA. cDNA coding for human gamma-1 constant region was cloned from a human cell line that secreted a human gamma-1 antibody by PCR. For convenient subcloning into the expression vector, BamHI sites were created at each end of the cDNA. In addition, a splice acceptor site and a 65 bp intron sequence were created at the 5'-end of the cDNA sequence. The BamHI fragment (1176 bp) containing the human gamma-1 cDNA splice acceptor site and intron sequence was substituted for the BamHI fragment (approximately 2.0 kb) in the existing heavy chain vector (Maeda et al., *supra*). The BamHI site to the 3'-side of the human gamma-1 constant region was then removed with Klenow polymerase.

2. Transfection of Expression Vectors

Expression vectors were introduced into Cos cells by electroporation using the Gene Pulsar apparatus (BioRad). DNA (10 μg of each vector) was added to a 0.8 ml aliquot of 1 x 10^7 cells/ml in PBS. A pulse was delivered at 1,900 volts, 25 μF capacitance. After a 10 min recovery period at ambient temperature, the electroporated cells were added to 8 ml of DMEM (GIBCO) containing 5% heat-inactivated gamma globulin-free fetal calf serum. After 72 h incubation, the medium was collected, centrifuged to remove cellular debris, and stored under sterile conditions at 4°C for short periods of time, or at -20°C for longer periods.

3. Purification of Humanized Antibodies

Supernatants from Cos cell transfections were pooled and purified on immobilized Protein A (ImmunoPure IgG Purification Kit, Pierce). The supernatant was sterilized by filtration through a 0.22 μm filter. After mixing with an equal volume of ImmunoPure IgG binding buffer (pH 8.0), the diluted sample was applied to a 1 ml protein A column and allowed to flow completely into the gel. After washing with 15 ml of ImmunoPure IgG binding buffer, the bound antibody was eluted with 5 ml of ImmunoPure IgG elution buffer (pH 2.8), and 1 ml fractions were collected. The pH of the first and second fractions was approximately 8.0. The pH of the third fraction was adjusted to physiological pH by the addition of 100 μl of ImmunoPure binding buffer. The five 1 ml fractions containing the Protein A-purified antibody were then assayed by ELISA to determine the amount of human IgG antibody present in each fraction. Antibody was detected using goat alkaline phosphate-conjugated anti-human IgG (whole molecule, Sigma).

4. Measurement of Binding Affinity

The binding of reshaped human 21.6 antibodies to $\alpha 4\beta 1$ integrin was assayed by ELISA in comparison with mouse and chimeric antibodies. Briefly, L cells transformed to express $\alpha 4\beta 1$ integrin on their cell surface were plated out and grown to confluence in 96-well tissue culture plates. The samples to be tested (either crude supernatants or protein-A-purified) were serially diluted and added to each well. After incubation for 1 h on ice and very gentle washing, goat anti-mouse or anti-human (gamma-chain specific) peroxidase conjugates (Sigma) were added. After a further 1 h incubation on ice and very gentle washing, the substrate (o-phenylenediamine dihydrochloride, Sigma) was added. After incubation for 30 min at room temperature, the reaction was stopped by adding 1 M H_2SO_4, and the A_{490} was measured.

Results from analyzing crude supernatants of the two versions of reshaped human 21.6 light chains (La and Lb), in combination with version Ha of reshaped human 21.6 heavy chain, indicated that the La version of reshaped human 21.6 V_L region

gave slightly better binding to antigen than version Lb. The La version was therefore used in subsequent experiments. Results from analysis of the crude supernatants of humanized 21.6 heavy chains (Ha and Hb), in combination with version La of humanized 21.6 light chain, showed no significant difference between the two versions (Ha and Hb) of reshaped human V_H regions. Version Ha was selected for use in further experiments because it contained only five changes in the human FRs compared with six changes in the human Hb.

Figure 12 compares binding of humanized 21.6 antibody (La + Ha) with chimeric 21.6 antibody. The data indicate that the reshaped human 21.6 antibody (La + Ha) bound to antigen as well as, and perhaps slightly better than, the chimeric 21.6 antibody. The chimeric 21.6 antibody is expected to be equivalent to mouse 21.6 antibody in its antigen binding characteristics because it contains the intact mouse 21.6 variable regions. The reshaped human 21.6 antibody (La + Ha) has also been shown to block binding to human $\alpha 4\beta 1$ integrin with an efficiency comparable to the original mouse 21.6 antibody and to the chimeric antibody. It is therefore concluded that reshaped human 21.6 antibody (La + Ha) has a specific binding affinity essentially equal to that of mouse 21.6 antibody. Moreover, because only minor modifications in the human FRs were necessary to recreate the antigen binding site of mouse 21.6 antibody within human variable regions, the reshaped human 21.6 antibody is predicted to behave like an authentic human antibody.

Reshaped human 21.6 antibody containing version La of the reshaped human 21.6 VL region and version Hc of the reshaped human 21.6 VH region was also tested for binding to L cells expressing human $\alpha 4\beta 1$ integrin on their surface in parallel with chimeric 21.6 antibody. The results indicate that reshaped human 21.6 antibody (La + Hc) binds well to antigen. The alteration in the CDR3 of the V_H region did not impair binding to antigen. Indeed, there is some indication that the alteration in the CDR3 may have slightly improved binding to antigen (Figure 12). Conceivably, the improvement may be more pronounced in a functional blocking assay.

Although the foregoing invention has been described in detail for purposes of clarity of understanding, it will be obvious that certain modifications may be practiced within the scope of the appended claims. All publications and patent documents cited above are hereby incorporated by reference in their entirety for all purposes to the same extent as if each were so individually denoted.

Table 4

Alignment of amino acid sequences leading to the design of reshaped human 21.6 light chain variable regions.

Kabat	#	FR or CDR	mouse 21.6	mouse kappa 5a	human kappa 1a	human REI	RH V_L 21.6	Comment
1	1	FR1	D	D	D	D	D	
2	2	|	I	I	I	I	I*	
3	3	|	Q	Q	Q	Q	Q	
4	4	|	M	M	M	M	M	
5	5	|	T	T	T	T	T	
6	6	|	Q	Q	Q	Q	Q	
7	7	|	S	S	S	S	S	
8	8	|	P	P	P	P	P	
9	9	|	S	S	S	S	S	
10	10	|	S	S	S	S	S	
11	11	|	L	L	L	L	L	
12	12	|	S	S	S	S	S	
13	13	|	A	A	A	A	A	
14	14	|	S	S	S	S	S	
15	15	|	L	L	V	V	V	
16	16	|	G	G	G	G	G	
17	17	|	G	D	D	D	D	
18	18	|	K	R	R	R	R	
19	19	|	V	V	V	V	V	
20	20	|	T	T	T	T	T	
21	21	|	I	I	I	I	I	
22	22	|	T	T	T	T	T	
23	23	FR1	C	C	C	C	C	
24	24	CDR1	K	R	R	Q	K	
25	25	|	T	A	A	A	T*	
26	26	|	S	S	S	S	S*	
27	27	|	Q	Q	Q	Q	Q*	
27A		|	-	D	S	-	-	
27B		|	-	-	L	-	-	
27C		|	-	-	V	-	-	
27D		|	-	-	X	-	-	

27E		|	-	-	X	-	-	
27F		|	-	-	-	-	-	
28	28	|	D	D	S	D	D*	
29	29	|	I	I	I	I	I*	
30	30	|	N	S	S	I	N*	
31	31	|	K	N	N	K	K*	
32	32	|	Y	Y	Y	Y	Y*	
33	33	|	M	L	L	L	M*	
34	34	CDR1	A	N	A	N	A	
35	35	FR2	W	W	W	W	W	
36	36	|	Y	Y	Y	Y	Y	
37	37	|	Q	Q	Q	Q	Q	
38	38	|	H	Q	Q	Q	Q	
39	39	|	K	K	K	T	T	K in CAMPATH-1H
40	40	|	P	P	P	P	P	
41	41	|	G	G	G	G	G	
42	42	|	K	G	K	K	K	
43	43	|	R	S	A	A	A	consider R in other versions
44	44	|	P	P	P	P	P	
45	45	|	R	K	K	K	R	supports L2 loop, consider K in other versions
46	46	|	L	L	L	L	L	
47	47	|	L	L	L	L	L	
48	48	|	I	I	I	I	I*	
49	49	FR2	H	Y	Y	Y	H	in middle of binding site, potential to interact with antigen, consider Y in other versions
50	50	CDR2	Y	Y	A	E	Y*	
51	51	|	T	A	A	A	T*	
52	52	|	S	S	S	S	S*	
53	53	|	A	R	S	N	A	
54	54	|	L	L	L	L	L	
55	55	|	Q	H	E	Q	Q	
56	56	CDR2	P	S	S	A	P	

57	57	FR3	G	G	G	G	G	
58	58	|	I	V	V	V	I	may be supporting L2, consider V in other versions
59	59	|	P	P	P	P	P	
60	60	|	S	S	S	S	S	
61	61	|	R	R	R	R	R	
62	62	|	F	F	F	F	F	
63	63	|	S	S	S	S	S	
64	64	|	G	G	G	G	G*	
65	65	|	S	S	S	S	S	
66	66	|	G	G	G	G	G	
67	67	|	S	S	S	S	S	
68	68	|	G	G	G	G	G	
69	69	|	R	T	T	T	R	adjacent to L1, on the surface near the binding site
70	70	|	D	D	D	D	D	
71	71	|	Y	Y	F	Y	Y*	F in CAMPATH-1H
72	72	|	S	S	T	T	T	
73	73	|	F	L	L	F	F	
74	74	|	N	T	T	T	T	
75	75	|	I	I	I	I	I	
76	76	|	S	S	S	S	S	
77	77	|	N	N	S	S	S	
78	78	|	L	L	L	L	L	
79	79	|	E	E	Q	Q	Q	
80	80	|	P	Q	P	P	P	
81	81	|	E	E	E	E	E	
82	82	|	D	D	D	D	D	
83	83	|	I	I	F	I	I	
84	84	|	A	A	A	A	A	
85	85	|	T	T	T	T	T	
86	86	|	Y	Y	Y	Y	Y	
87	87	|	Y	F	Y	Y	Y	
88	88	FR3	C	C	C	C	C	

89	89	CDR3	L	Q	Q	Q	L	
90	90	|	Q	Q	Q	Q	Q*	
91	91	|	Y	G	Y	Y	Y*	
92	92	|	D	N	N	Q	D*	
93	93	|	N	T	S	S	N*	
94	94	|	L	L	L	L	L*	
95		|	-	P	P	P	-	
95A		|	-	P	E	-	-	
95B		|	-	-	-	-	-	
95C		|	-	-	-	-	-	
95D		|	-	-	-	-	-	
95E		|	-	-	-	-	-	
95F		|	-	-	-	-	-	
96	95	|	W	R	W	Y	W*	
97	96	CDR3	T	T	T	T	T	
98	97	FR4	F	F	F	F	F	
99	98	|	G	G	G	G	G	
100	99	|	G	G	Q	Q	Q	
101	100	|	G	G	G	G	G	
102	101	|	T	T	T	T	T	
103	102	|	K	K	K	K	K	
104	103	|	L	L	V	L	V	as in CAMPATH-1H
105	104	|	E	E	E	Q	E	as in CAMPATH-1H
106	105	|	I	I	I	I	I	
106A		|	-	-	-	-	-	
107	106	FR4	K	K	K	T	K	as in CAMPATH-1H

Legend: (Kabat) numbering according to Kabat et al., *supra*; (#) sequential numbering as used in the molecular modelling; (mouse 21.6) amino acid sequence of the V_L region from mouse 21.6 antibody; (mouse kappa 5) consensus sequence of mouse kappa V_L regions from subgroup 5 (Kabat et al., *supra)*; (human kappa 1) consensus sequence of human V_L regions from subgroup 1 (Kabat et al., *supra)*; (human REI) amino acid sequence of a human V_L region (Palm et al. (1975), *supra*); (RH V_L 21.6) amino acid sequence of version L1 of reshaped human 21.6 V_L region; (*) residues that are part of the canonical structures for the CDR loops (Chothia et al., *supra)*; (underlined) residues in the human FRs where the amino acid residue was changed.

Table 5

Alignment of amino acid sequences leading to the design of reshaped human 21.6 heavy chain variable regions.

Kabat	#	FR or CDR	mouse 21.6	mouse 2c	human 1	human 21/28'CL	RH V_H 21.6	Comment
1	1	FR1	E	E	Q	Q	Q	
2	2	|	V	V	V	V	V	
3	3	|	Q	Q	Q	Q	Q	
4	4	|	L	L	L	L	L	
5	5	|	Q	Q	V	V	V	
6	6	|	Q	Q	Q	Q	Q	
7	7	|	S	S	S	S	S	
8	8	|	G	G	G	G	G	
9	9	|	A	A	A	A	A	
10	10	|	E	E	E	E	E	
11	11	|	L	L	V	V	V	
12	12	|	V	V	K	K	K	
13	13	|	K	K	K	K	K	
14	14	|	P	P	P	P	P	
15	15	|	G	G	G	G	G	
16	16	|	A	A	A	A	A	
17	17	|	S	S	S	S	S	
18	18	|	V	V	V	V	V	
19	19	|	K	K	K	K	K	
20	20	|	L	L	V	V	V	
21	21	|	S	S	S	S	S	
22	22	|	C	C	C	C	C	
23	23	|	T	T	K	K	K	
24	24	|	A	A	A	A	A	
25	25	|	S	S	S	S	S	
26	26	|	G	G	G	G	G*	
27	27	|	F	F	Y	Y	F*	H1 canonical structure, consider Y in other versions
28	28	|	N	N	T	T	N*	H1 canonical structure, on the surface

29	29	|	I	I	F	F	I*	H1 canonical structure, consider F in other versions
30	30	FR1	K	K	T	T	K*	H1 canonical structure, on the surface
31	31	CDR1	D	D	S	S	D*	
32	32	|	T	T	Y	Y	T*	
33	33	|	Y	Y	A	A	Y	
34	34	|	I	M	I	M	I*	
35	35	|	H	H	S	H	H	
35A		|	-	-	-	-	-	
35B		CDR1	-	-	-	-	-	
36	36	FR2	C	W	W	W	W	buried residue, no obvious special role for C
37	37	|	V	V	V	V	V	
38	38	|	K	K	R	R	R	
39	39	|	Q	Q	Q	Q	Q	
40	40	|	R	R	A	A	A	
41	41	|	P	P	P	P	P	
42	42	|	E	E	G	G	G	
43	43	|	Q	Q	Q	Q	Q	
44	44	|	G	G	G	R	R	V_L-V_H packing, consider G in other versions
45	45	|	L	L	L	L	L	
46	46	|	E	E	E	E	E	
47	47	|	W	W	W	W	W	
48	48	|	I	I	M	M	M	
49	49	FR2	G	G	G	G	G	
50	50	CDR2	R	R	W	W	R	
51	51	|	I	I	I	I	I	
52	52	|	D	D	N	N	D	
52A	53	|	P	P	P	A	P*	
52B		|	-	-	Y	-	-	
52C		|	-	-	-	-	-	
53	54	|	A	A	G	G	A*	

54	55	|	N	N	N	N	N*	
55	56	|	G	G	G	G	G*	
56	57	|	Y	N	D	N	Y	
57	58	|	T	T	T	T	T	
58	59	|	K	K	N	K	K	
59	60	|	Y	Y	Y	Y	Y	
60	61	|	D	D	A	S	D	
61	62	|	P	P	Q	Q	P	
62	63	|	K	K	K	K	K	
63	64	|	F	F	F	F	F	
64	65	|	Q	Q	Q	Q	Q	
65	66	CDR2	G	G	G	G	G	
66	67	FR3	K	K	R	R	R	
67	68	|	A	A	V	V	V	
68	69	|	T	T	T	T	T	
69	70	|	I	I	I	I	I	
70	71	|	T	T	T	T	T	
71	72	|	A	A	A	R	A*	H2 canonical structure, supporting H2
72	73	|	D	D	D	D	D	
73	74	|	T	T	T	T	T	
74	75	|	S	S	S	S	S	
75	76	|	S	S	T	A	A	
76	77	|	N	N	S	S	S	
77	78	|	T	T	T	T	T	
78	70	|	A	A	A	A	A	
79	80	|	Y	Y	Y	Y	Y	
80	81	|	L	L	M	M	M	
81	82	|	Q	Q	E	E	E	
82	83	|	L	L	L	L	L	
82A	84	|	S	S	S	S	S	
82B	85	|	S	S	S	S	S	
82C	86	|	L	L	L	L	L	
83	87	|	T	T	R	R	R	
84	88	|	S	S	S	S	S	
85	89	|	E	E	E	E	E	
86	90	|	D	D	D	D	D	

87	91	|	T	T	T	T	T	
88	92	|	A	A	A	A	A	
89	93	|	V	V	V	V	V	
90	94	|	Y	Y	Y	Y	Y	
91	95	|	F	Y	Y	Y	Y	
92	96	|	C	C	C	C	C	
93	97	|	A	A	A	A	A	
94	98	FR3	R	R	R	R	R*	
95	99	CDR3	E	G	A	G	E	
96	100	|	G	Y	P	G	G	
97	101	|	Y	Y	G	Y	Y	
98	102	|	Y	Y	Y	Y	Y	
99	103	|	G	Y	G	G	G	
100	104	|	N	D	S	S	N	
100A	105	|	Y	S	G	G	Y	
100B	106	|	G	X	G	S	G	
100C	107	|	V	V	G	-	V	
100D	108	|	Y	G	C	-	Y	
100E	109	|	A	Y	Y	-	A	
100F	110	|	M	Y	R	-	M	
100G		|	-	A	G	-	-	
100H		|	-	M	D	-	-	
100I		|	-	-	Y	-	-	
100J		|	-	-	X	-	-	
100K		|	-	-	F	-	-	
101	111	|	D	D	D	N	D	
102	112	CDR3	Y	Y	Y	Y	Y	
103	113	FR4	W	W	W	W	W	
104	114	|	G	G	G	G	G	
105	115	|	Q	Q	Q	Q	Q	
106	116	|	G	G	G	G	G	
107	117	|	T	T	T	T	T	
108	118	|	S	X	L	L	L	
109	119	|	V	V	V	V	V	
110	120	|	T	T	T	T	T	
111	121	|	V	V	V	V	V	
112	122	|	S	S	S	S	S	
113	123	FR4	S	S	S	S	S	

Legend: (Kabat) numbering according to Kabat et al., *supra*; (#) sequential numbering as used in the molecular modelling; (mouse 21.6) amino acid sequence of the V_H region from mouse 21.6 antibody; (mouse 2c) consensus sequence of mouse V_H regions from subgroup 2c (Kabat et al., *supra)*; (human 1) consensus sequence of human V_H regions from subgroup 1 (Kabat et al., *supra)*; (human 21/28'CL) amino acid sequence of a human V_H region (Dersimonian et al. (1987), *supra)*; (RH V_H 21.6) amino acid sequence of version H1 of reshaped human 21.6 V_H region; (*) residues that are part of the canonical structures for the CDR loops (Chothia et al., *supra*); (underlined) residues in the human FRs where the amino acid residue was changed.